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# M. LÕHMUS, Anne PAJU, N. SAMEL, M. LOPP, Ü. LILLE

# ADSORPTION HIGH-PERFORMANCE LIQUID CHROMATOGRAPHY OF PROSTANOIDS, USE OF WATER-CONTAINING MOBILE PHASES FOR SEPARATION OF 15R/S-ISOMERS OF CLOPROSTENOL, AND PROSTAGLANDINS E<sub>1</sub> AND E<sub>2</sub>

Reversed-phase high-performance liquid chromatography (HPLC) has proved a very useful tool for separating prostaglandins (PG) and their analogs [1-4]. Nevertheless, for the resolution of many PG-isomers the hydrophobic stationary phase offers no sufficient selectivity. If silica gel is used for HPLC resolutions, derivation of the acid moiety is necessary prior to chromatography [5-7]. Moreover, acidic modifiers in the mobile phase or silicaic acid columns are used in the separation of nonderived PGs to avoid peak tailing and to improve resolution efficiency [8, e]. These methods hardly meet the requirements for preparative-scale separations due to the difficulty in recovering pure initial components and to the instability of some PGs (e.g. the E-series) under acidic conditions [10].

The improvement in the resolution by deactivating strong adsorption sites with water as a mobile phase component has been shown by several authors [ $^{11-14}$ ], but the separation of carboxylic acids as well as PGs has not yet been performed by this method. We demonstrate here the improvement in the separation efficiency using a trace amount of water as a mobile phase component for several solvent systems in separating 17,18,19,20-tetranor-16(3-chloro)-phenoxy-PGF<sub>2α</sub> (cloprostenol (2)) from its 15*R*-isomer (1) and PGE<sub>1</sub> (4) from PGE<sub>2</sub> (3) (Fig. 1).





(1)





17,18,19,20-tetranor-16-(3-chloro)-phenoxy-PGF<sub>2¢</sub> (cloprostenol)





Fig. 1. Structures of the compounds studied.

A Du Pont HPLC system N 8845 with UV-spectrophotometrical and refractive index (RI) detectors was used. The Zorbax SIL column ( $150 \times \times 4.6 \text{ mm I.D.}$ ) theoretical plate count was 9700. The column compartment operated at 35 °C. The eluent flow rate was 0.6 ml/min. 15R/S-isomers of cloprostenol were detected spectrophotometrically at 269 or 280 nm (when the solvent system contained benzene) or refractometrically (when the solvent system contained acetone). PGE<sub>1</sub> and PGE<sub>2</sub> were detected refractometrically and spectrophotometrically at 211 nm (when hexane—isopropanol—water was used). The two-pen strip chart recorder operated at a speed of 10 or 20 cm/h.

**Solvents**. All solvents (analytical grade) were purchased from Reakhim (USSR). Hexane, ethanol and 1,4-dioxane were used without purification. Methanol was redistilled. Benzene and acetone were redistilled over  $P_2O_5$ . Chloroform was washed with  $H_2O$ , dried over  $CaCl_2$  and redistilled over  $P_2O_5$ . Ethyl acetate was washed with 5% aqueous  $Na_2CO_3$  and saturated  $CaCl_2$ , dried over  $K_2CO_3$  and redistilled over  $P_2O_5$ . Acetonitrile and isopropanol were rectified. Bidistilled water was used. The water content in the solvents was checked on a Chrom-5 gas chromatograph equipped with a katharometer. 1.5 m glass columns with Teparon-18 and Separon CHN stationary phases were used. The solvent responses were calibrated vs. water response. The water content in ethanol (4.2% v), acetone (0.2% v) and dioxane (1.2% v) was taken into consideration in calculating solvent composition. All the other solvents contained less than 0.1% of water before mixing. The water content in the mobile phases varied from 0 to 100% of its saturation at ambient temperature. In all cases, the ternary eluting solutions used were transparent and there were no water droplets on the reservoir walls. The columns were equilibrated by pumping 20 ml of each eluting mixture through column before injections.

**Samples.** Cloprostenol was synthesized by the authors and identified by <sup>13</sup>C NMR spectroscopy.\* Biosynthetical natural prostaglandins  $E_1$  and  $E_2$  were purchased from the Pilot Production Plant of Organic Synthesis and Biopreparations (Institute of Chemistry, Tallinn). Samples were dissolved in chloroform at a concentration of 10–20 mg/ml. When UV-detection was used, a 20–100 µg sample was injected, in case of RI-detection 100–500 µg.

**Calculations.** The capacity factors (k'), resolution factors  $(\alpha)$ , plate counts  $(N_{5\sigma})$  and peak asymmetry factors were calculated, and the resolution function  $(R_s)$  was estimated according to L. R. Snyder and J. J. Kirkland[<sup>14</sup>]. The parameters were the arithmetic means of three measurements. The column void volume was measured as elution volume of toluene by using the eluting system hexane—isopropanol=75/25 (v/v) which was found to be 1.89 ml.

### Results and discussion

It is well known that the 15 R- and S-isomers of prostaglandins are preparatively separable on silica gel<sup>[15]</sup>. We have found, however, that the use of carefully dried solvents leads to very broad and unresolved peaks of these compounds on the Zorbax SIL column (Fig. 2). If water was added to the eluting mixture, the peak shape improved drastically, and the column efficiency increased (Table 1). The best resolution was

\* The authors are indebted to T. Välimäe for interpreting the <sup>13</sup>C NMR data.

Fig. 2. Separation of cloprostenol from its 15*R*-isomer. Column Zorbax SIL (4.6×150 mm); mobile phase — hexane—isopropanol = 80:20 v/v; flow rate — 0.6 ml/min; UV-spectrophotometer — 269 nm; absorbance — 0.08 AUFS; column temperature — 35 °C; ~ 100 μg injected; peaks, see Fig. 1.

Fig. 3. Separation of cloprostenol from its 15*R*-isomer. Column Zorbax SIL ( $4.6 \times 150$  mm); mobile phase — hexane—isopropanol—water = 85:14.25:0.75v/v/v; flow rate — 0.6ml/min; UV-sepctrophotometer — 269 nm; absorbance — 0.08 AUFS; column temperature —  $35^{\circ}$ ; ~20 µg injected; peaks, see Fig. 1.



achieved with a maximum water content in the eluting mixture (Fig. 3), and therefore it was subsequently used in each case. It is notable that benzene as a less polar component of the mobile phase offers a better resolution of compounds (1) and (2) than hexane (Table 2). When comparing different ternary eluting systems it can be seen that benzene acetonitrile—water and benzene—acetone—water provide the highest selectivity in resolution, but the former (Fig. 4) is more advantageous due to UV transmission problems. It can also be supposed that methanol

# Dependence of peak asymmetry factor and plate count on the water content in hexane—isopropanol and benzene—isopropanol mixtures

Eluent composition, v/v/v	k'	Peak asym- metry factor	$N_{5\sigma}$
Hexane—isopropanol—water	10.0		00
80:20 :0	10.0	4.0	45
80:19.8 :0.2	5.2	2.5	65
80:19.6 :0.4	4.0	1.9	250
80:19.4 :0.6	3.6	1.3	850
80:19.2 :0.8	3.5	1.1	1800
80:19 :1.0	3.5	1.0	4200
85:14.25:0.75	8.1	1.0	4100
Benzene-isopropanol-water			
90:10 :0	12.7	4.0	20
90: 9.95:0.05	9.4	3.8	60
90: 9.9 :0.1	7.0	3.5	150
90: 9.8 .0.2	5.0	3.0	580
90: 9.6 :0.4	4.4	2.3	1000
90: 9.5 :0.5	3.8	1.3	2400
95: 4.8 :0.2	3.1	2.0	1250
80:18.6:1.4	1.2	1.3	0900

Sample: 17,18,19,20-tetranor-16(3-chloro)-phenoxy-PGF2a

Table 2

## Mobile phase selection for resolving 15S- and 15R-isomers of 17,18,19,20-tetranor-16(3-chloro)-phenoxy-PGF<sub>2n</sub>

k' values are given for 15S-isomer; a values are expressed as the ratio of k' value of 15S-isomer to k' of 15R-isomer;  $N_{5\sigma}$  and the peak asymmetry factor are arithmetic means of both isomers

Eluent composition, v/v/v	k'	α	Peaks asym- metry factor	N <sub>58</sub>
Hexane isopropagal water				
80 :19 :1	3.5	1.13	1	4200
Benzene-isopropanol-water	. Salar It.	A starting	The stades	
90 : 9.5 :0.5 Benzene ethanol water	3.8	, 1.36	1.3	2400
93 : 6.64:0.36	5.9	1.35	1	6000
Benzene-methanol-water	at left produ	lender Maria	way and	
95.5: 4.27:0.23	9.6	1.31	1	9800
60 :39.2:0.8	8.8	1.20	I	6200
Benzene-dioxane-water			and the second	
70 :29.32:0.68	4.1	1.42	1.4	2000
70 :29.40:0.60	5.5	1.48	1	5000
Benzene-acetonitrile-water				
60 :38.8 :1.2 Bonzono postonitrilo	4.5	1.48	10 10 10 10	6800
60 :40	12.8	1.51	4.0	300
Benzene-acetonitrile-methanol	designine 34	15	Participation (1991)	1500
60 :38.8 :1.2	5.5	1.54	2.0	1500

and ethanol support the blocking of strong adsorption sites of silica gel by water, and therefore they increase the plate count (Table 2).

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Fig. 4. Separation of cloprostenol from its 15*R*-isomer. Column Zorbax SIL (4.6 $\times$ 150 mm); mobile phase — benzene—acetonitrile—water = 60:38.8:1.2 v/v/v; flow rate— 0.6 ml/min; UV-spectrophotometer — 280 nm; absorbance — 0.16 AUFS; column temperature — 35°; ~30 µg injected; peaks, see Fig. 1.

Fig. 5. Separation of PGE<sub>2</sub> from PGE<sub>1</sub>. Column Zorbax SIL ( $4.6 \times 150 \text{ mm}$ ); mobile phase — hexane—isopropanol—water = 90:9.6:0.4 v/v/v; flow rate — 0.6 ml/min; UV-spectrophotometer — 211 nm; absorbance — 0.64 AUFS; RI-detector —  $0.05 \times 10^3$  RI units full scale; column temperature — 35°; 310 µg of PGE<sub>2</sub> and 250 µg of PGE<sub>1</sub> injected; peaks, see Fig. 1.

Data about selecting mobile phases for resolution of PGE1 and PGE2 k' is given for PGE<sub>1</sub>;  $\alpha$  is expressed as the ratio the k' value of PGE<sub>1</sub> to the k' value of PGE<sub>2</sub>

Eluent composition, v/v/v	k'	antes a contraction	Rs
Banzana mathanal water	demidation of E	Pullyde R. R. Dy	
95.5: 4.32:0.18	8.7	1.21	large
Benzene—ethanol—water	83	1.20	large
Benzene—isopropanol—water	0.0	1.20	Inge
92 : 7.68:0.32 Hexane_isopropanol_water	4.2	1.18	1.25
92 : 7.7 :0.3	8.6	1.17	large
Chloroform—methanol—water	7.8	1.11	0.95
Hexane—ethylacetate—water	1.0	Sura Tittues	0.50
40 : 5.94:0.6	6.6	and no 1.11 3 (j	0.95
80 :19.65:0.35	9.5	1.09	0.8
Chloroform—ethanol—water	E A	1.07	0.8
Chloroform—isopropanol—water	0.4	1.07	0.0
92 : 7.72:0.28	4.8	1.06	0.7
65 :34.2 :0.8	4.3	1.05	0.6
Hexane—dioxane—water	0.4	1.05	0.7
Chloroform—acetone—water	9.4	1.05	0.7
80 :19.57:0.43	12.5	1.04	07.5
65 :33.95:1.05	4.5	~1.0	
Chloroform—ethylacetate—water	5.0	.10	
Benzene—dioxane—water	0.3	~1.0	
80 :19.76:0.24	4.8	~1.0	
80 :19.74:0.26	6.5	0.96	0.6

Selecting a suitable mobile phase for the resolution of PGE<sub>1</sub> and  $PGE_2$  as free acids, we found that the eluting mixtures containing solvents of extremely different polarity yield the best selectivity in resolution (Table 3). It is noteworthy that a relatively high column load (Fig. 5) may be achieved with no significant decrease in column efficiency. This enables to resolve preparatively the above-mentioned compounds.

Plate counts  $N_{5\sigma}$  for PGE<sub>1</sub> and PGE<sub>2</sub> were on the average 1.5 times lower than for R/S-isomers of cloprostenol. This was probably due to the lower water content in the mobile phase capable of producing acceptable k' values in the PGE-series (a strong solvent content in the eluting mixture should be lower than for the PGF-series).

In conclusion it may be said that further investigation is needed to elucidate the conditions of the water-deactivation of silica gels in resolving different prostaglandins and their analogs.

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Academy of Sciences of the Estonian SSR, Institute of Chemistry

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#### M. LÖHMUS, Anne PAJU, N. SAMEL, M. LOPP, U. LILLE

# PROSTANOIDIDE KÖRGEFEKTIIVNE VEDELIKU-ADSORPTSIOON-**KROMATOGRAAFIA. VETT SISALDAVATE LIIKUVATE FAASIDE** KASUTAMINE KLOPROSTENOOLI 15R/S-ISOMEERIDE JA PROSTAGLANDIINIDE E1 JA E2 LAHUTAMISEKS

On näidatud kolonni efektiivsuse tõus ja piigi sümmeetrilisuse paranemine vee sisalduse kasvamisel solventsüsteemides heksaan—isopropanool ja benseen—isopropanool. Määrati 8 ja 16 vett sisaldava solventsüsteemi selektiivsus vastavalt kloprostenooli 15*R*- ja 15 *S*isomeeride lahutamisel ja prostaglandiinide E1 ja E2 lahutamisel.

#### М. ЛЫХМУС, Анне ПАЮ, Н. САМЕЛЬ, М. ЛОПП, Ю. ЛИЛЛЕ

# АДСОРБЦИОННАЯ ВЫСОКОЭФФЕКТИВНАЯ ЖИДКОСТНАЯ ХРОМАТОГРАФИЯ ПРОСТАНОИДОВ И ИСПОЛЬЗОВАНИЕ ВОДОСОДЕРЖАЩИХ ПОДВИЖНЫХ ФАЗ ДЛЯ РАЗДЕЛЕНИЯ 15R/S-ИЗОМЕРОВ КЛОПРОСТЕНОЛА И ПРОСТАГЛАНДИНОВ Е1 И Е2

Показано, что с повышением содержания воды в сольвентных системах гексан-изопропанол и бензол-изопропанол возрастают эффективность колонки и четкость разделения пиков. Определена селективность 8 и 16 водосодержащих сольвентных систем для разделения 15R/S-изомеров клопростенола и простагландинов E1 и E2 соответственно.