

## AN ADVANCED INTERMEDIATE FOR THE SYNTHESIS OF 9,11-SECOSTEROLS

Raissa JÄÄLAID<sup>a</sup>, Ivar JÄRVING<sup>a</sup>, Tõnis PEHK<sup>b</sup>, and Ülo LILLE<sup>a</sup>

<sup>a</sup> Institute of Chemistry at Tallinn Technical University, Akadeemia tee 15, EE-0026 Tallinn, Estonia, e-mail: lille@boc.ic.ee

<sup>b</sup> Institute of Chemical and Biological Physics, Akadeemia tee 23, EE-0026 Tallinn, Estonia

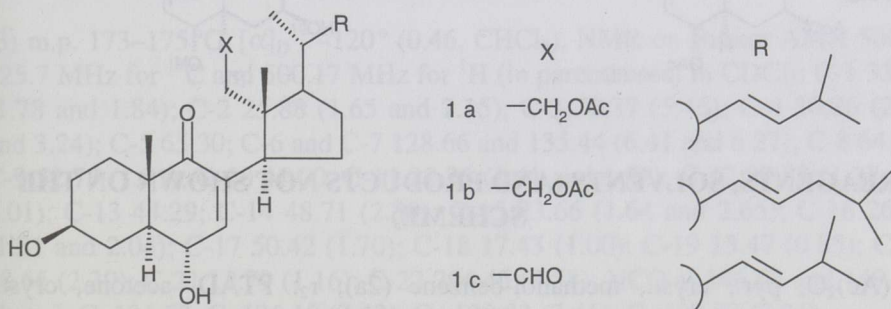
Received 5 December 1997

SOBIVALT FUNKTSIONALISEERITUD VAHEÜHEND 9,11-SEKOSTEROOLIDE SÜNTEESIKS. Raissa JÄÄLAID, Ivar JÄRVING, Tõnis PEHK ja Ülo LILLE

**Key words:** partial synthesis of sterols, modification of B- and C-rings in sterols.

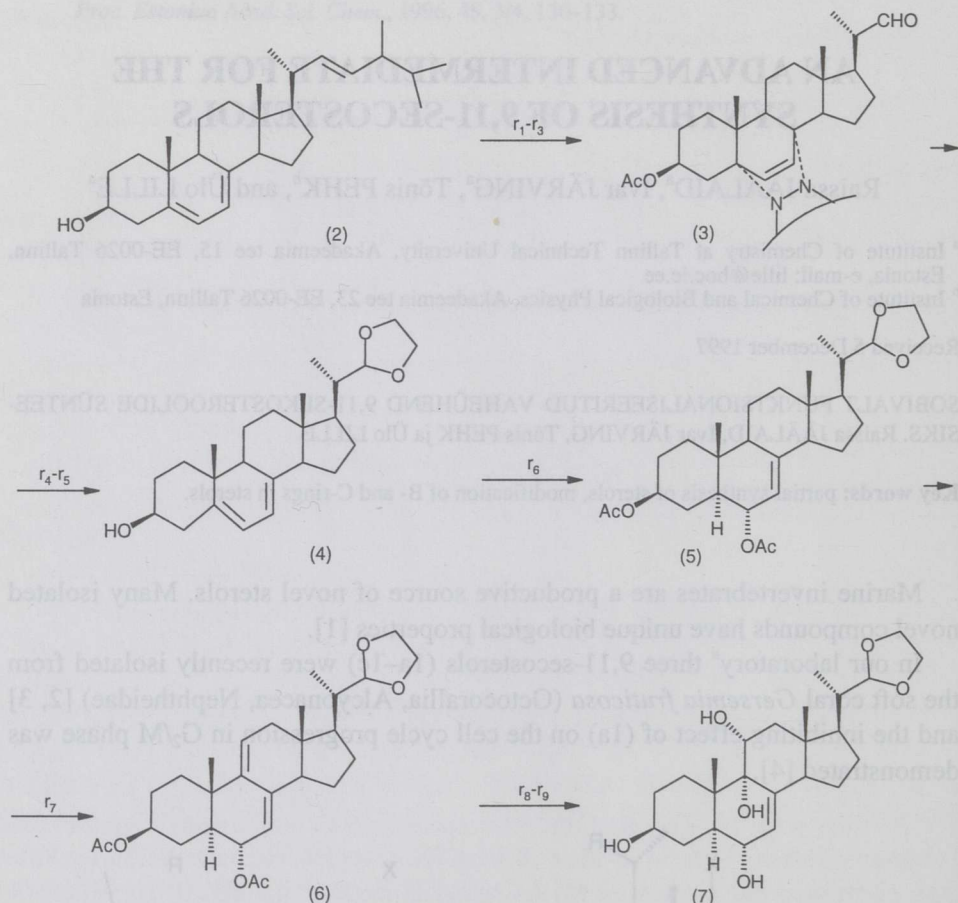
Marine invertebrates are a productive source of novel sterols. Many isolated novel compounds have unique biological properties [1].

In our laboratory<sup>a</sup> three 9,11-secosterols (1a–1c) were recently isolated from the soft coral *Gersemia fruticosa* (Octocorallia, Alcyonacea, Nephtheidae) [2, 3] and the inhibiting effect of (1a) on the cell cycle progression in G<sub>2</sub>/M phase was demonstrated [4].



In this communication we propose an advanced intermediate for partial synthesis of 9,11-secosterols. For some earlier examples of partial synthesis of sterols see in [5, 6].

We started with cheap and stereochemically proper ergosterol (2) and the transformations shown on the following scheme were carried out.



### REAGENTS, SOLVENTS (AND PRODUCTS NOT SHOWN ON THE SCHEME)

$r_1$ :  $(Ac)_2O$ , pyr., cryst., methanol-benzene (2a);  $r_2$ : PTAD, acetone, cryst., ethanol-benzene (2b);  $r_3$ : ozone, pyr.,  $CH_2Cl_2$ ,  $-78^\circ C$  [7], chrom., benzene-acetone /20:1/;  $r_4$ :  $CH_2OHCH_2OH$ , pTSA,  $CH_2Cl_2$ , chrom., petrol ether-acetone /8:1-6:1/(3a);  $r_5$ :  $LiAlH_4$ , THF,  $\Delta$ , chrom., chloroform-acetone /50:1-40:1/,  $r_1$ : (4a);  $r_6$ : a.  $BH_3$ , THF, b.  $H_2O_2$ , NaOH, c.  $r_1$ , chrom., petrol ether-acetone-

dichloromethane /15:1:1/; r<sub>7</sub>: Hg(OCOCH<sub>3</sub>)<sub>2</sub>, CHCl<sub>3</sub>, CH<sub>3</sub>COOH, chrom., petrol ether-diethylether /5:1/ (6a–6b); r<sub>8</sub>: OsO<sub>4</sub>, dioxane, chrom., chloroform-ethanol /9:1/ (6c); r<sub>9</sub>: LiAlH<sub>4</sub>, diethylether, chrom., chloroform-ethanol /9:1/ (r<sub>6</sub>–r<sub>8</sub> as in [5]). (2) from "Sigma", min. 90% purity.

All reaction products were purified over silica (if not stated otherwise) using flash chromatography under slight pressure and were obtained as crystalline substances. The yields were not optimized.

The aldehyde (3) was obtained as pure 20S epimer,  $[\alpha]_D^{19} -120^\circ$  (0.46, CHCl<sub>3</sub>). After customary protection/deprotection procedures, resulting in (4), ethylene acetal (5) was prepared smoothly with the necessary  $\alpha$ -orientated (equatorial) hydroxyl group at C-6 (9.8 Hz coupling between H-6 and H-5). In the dehydrogenation step three chromatographic fractions were obtained. The expected diene-acetal (6) was found in the second fraction together with the starting acetal (5). In the first and third fractions the corresponding 3-acetyl- and 3,6,12-triacetyldiene acetals were identified/designated respectively as (6a) and (6b) [8]. The intermediate osmate ester (6c) was cleaved immediately to the tetrolacetal (7) obtained from ergosterol with ca 10% yield.

The 3 $\beta$ ,6 $\alpha$ ,9 $\alpha$ ,11 $\alpha$ -tetrahydroxy-5 $\alpha$ -pregn-7-ene-20(S)-carbaldehyde ethylene acetal (7) was prepared using quite a short sequence of transformations, whose preliminary outlines were given above. It is an advanced intermediate in the synthesis towards target compounds. This molecule has necessary functionalities for introduction of the side chain and for selective cleavage of the bond between carbon atoms C-9 and C-11 resulting in keto functions at these atoms. With the aim of improving the limiting dehydrogenation step, other methods, among them microbiological 11-hydroxylation of ethylene acetal (5), are being studied.

## CHARACTERIZATION OF COMPOUNDS

(2a) m.p. 177–179, (2b) 173–174°C.

(3) m.p. 173–175°C,  $[\alpha]_D^{19} -120^\circ$  (0.46, CHCl<sub>3</sub>), NMR on Bruker AMX 500 at 125.7 MHz for <sup>13</sup>C and 500.17 MHz for <sup>1</sup>H (in parentheses) in CDCl<sub>3</sub>: C-1 33.62 (1.78 and 1.84); C-2 25.88 (1.65 and 2.15); C-3 70.37 (5.46); C-4 30.86 (2.21 and 3.24); C-5 65.30; C-6 and C-7 128.66 and 135.44 (6.41 and 6.27); C-8 64.58; C-9 52.73 (1.81); C-10 41.00; C-11 22.26 (1.41 and 1.51); C-12 37.83 (1.37 and 2.01); C-13 44.29; C-14 48.71 (2.39); C-15 23.66 (1.64 and 2.65); C-16 26.00 (1.51 and 2.04); C-17 50.42 (1.70); C-18 17.43 (1.00); C-19 13.47 (0.85); C-20 48.66 (2.39); C-21 13.79 (1.16); C-22 204.45 (9.58); NCO at 146.68 and 149.12; phenyl: C<sub>s</sub> 131.53; C<sub>o</sub> 126.18 (7.42); C<sub>m</sub> 128.83 (7.41); C<sub>p</sub> 127.83 (7.31).

(3a) m.p. 204–206°C.

(4) m.p. 163–165°C.



(4a) m.p. 144–147°C NMR (in CDCl<sub>3</sub>): C-1 37.87 (1.36 and 1.89); C-2 28.08 (1.58 and 1.91); C-3 72.75 (4.69); C-4 36.63 (2.38 and 2.47); C-5 141.16; C-6 and C-7 120.14 and 116.47 (6.22 and 6.45); C-8 138.68; C-9 46.01 (2.00); C-10 37.07; C-11 23.17 (1.44 and 1.73); C-12 38.93 (1.31 and 2.08); C-13 43.26; C-14 53.82 (1.91); C-15 20.99 (2.04); C-16 27.25 (1.45 and 2.00); C-17 52.05 (1.51); C-18 11.63 (0.63); C-19 16.14 (0.95); C-20 39.60 (1.81); C-21 11.71 (0.95); C-22 106.01 (4.87); —O—CH<sub>2</sub>—CH<sub>2</sub>—O— 65.04 and 65.21 (3.86, 3.87, 3.93, 3.95); OAc 21.36 (2.04) and 170.45.

(5) m.p. 205–207°C,  $[\alpha]_D^{23} +76^\circ$  (0.26, CHCl<sub>3</sub>), (5) NMR (in CDCl<sub>3</sub>): C-1 36.63 (1.24 and 1.81); C-2 27.11 (1.49 and 1.83); C-3 72.62 (4.68); C-4 29.63 (1.34 and 1.93); C-5 44.80 (1.58); C-6 73.06 (5.06); C-7 118.03 (5.05); C-8 142.68; C-9 48.76 (1.78); C-10 35.30; C-11 22.83 (1.43 and 1.61); C-12 38.93 (1.31 and 2.04); C-13 43.89; C-14 54.13 (1.87); C-15 21.22 (1.47 and 1.63); C-16 26.97 (1.45 and 1.96); C-17 52.20 (1.51); C-18 11.75 (0.55); C-19 13.83 (0.92); C-20 39.47 (1.78); C-21 11.66 (0.94); C-22 105.88 (4.85); —O—CH<sub>2</sub>—CH<sub>2</sub>—O— 65.02 (3.85 and 3.97) and 65.18 (3.85 and 3.93); OAc 21.31 (2.05) and 21.36 (2.03); 170.54 and 171.26.

(6) MS: 472 (M<sup>+</sup>), 412 (M<sup>+</sup>–60), 352 (M<sup>+</sup>–120), UV,  $\lambda_{\max}$ : 251.5, 242.0, 235.0 nm.

(6a) MS: 414 (M<sup>+</sup>), 354 (M<sup>+</sup>–60), UV,  $\lambda_{\max}$ : 250.5 nm.

(6b) MS: 531 (M<sup>+</sup>+1), 471 (M<sup>+</sup>–59), 410 (M<sup>+</sup>–120), 350 (M<sup>+</sup>–180), UV,  $\lambda_{\max}$ : 249.5, 242.5, 236.0 nm.

(7) m.p. 212–213°C,  $[\alpha]_D^{18} +10$  (0.70, CHCl<sub>3</sub>), NMR (in pyridine d<sub>5</sub>): C-1 33.05 (2.46a, 2.82e); C-2 32.70 (2.02a, 2.32e); C-3 70.24 (4.11a); C-4 36.00 (3.31e, 1.97a); C-5 43.51 (2.66); C-6 69.56 (4.32); C-7 129.56 (5.99); C-8 139.57; C-9 75.18; C-10 41.25; C-11 69.53 (4.60); C-12 47.41 (2.56e, 2.14a); C-13 43.64; C-14 50.63 (2.91); C-15 23.46 (1.61); C-16 27.43 (1.56 and 2.06); C-17 52.60 (1.90); C-18 12.47 (0.80); C-19 16.21 (1.44); C-20 39.96 (2.01); C-21 12.29 (1.24); C-22 106.10 (5.02); —O—CH<sub>2</sub>—CH<sub>2</sub>—O— 65.58 and 65.66 (3.86, 3.89, 3.98 and 4.00).

NMR data were assigned by 2D <sup>1</sup>H - <sup>1</sup>H and <sup>1</sup>H - <sup>13</sup>C COSY correlation diagrams. MS measurements: EI – 70 eV, CI – isobutane, gas-chromatographic separation: 200–300°C, RLS –150, 10 m.

## ACKNOWLEDGEMENTS

The authors are grateful to M. Müürisepp and M. Liiv for the MS analyses and to the Estonian Science Foundation for the financial support (grant No. 2088).

## REFERENCES

1. D'Auria, M. V., Minale, M. & Riccio, R. Polyoxygenated steroids of marine origin. *Chem. Rev.*, 1993, **93**, 1839–1895.
2. Koljak, R., Pehk, T., Järving, I., Liiv, M., Lopp, A., Varvas, K., Vahemets, A., Lille, Ü. & Samel, N. New antiproliferative 9,11-secoesterol from soft coral *Gersemia fruticosa*. *Tetrahedron Lett.*, 1993, **34**, 12, 1985–1986.
3. Koljak, R., Lopp, A., Pehk, T., Varvas, K., Müürisepp, M., Järving, I. & Samel, N. New cytotoxic sterols from the soft coral *Gersemia fruticosa*. *Tetrahedron*, 1998, **54**, 179–186.
4. Lopp, A., Pihlak, A., Paves, H., Samuel, K., Koljak, R. & Samel, N. The effect of 9,11-secoesterol, a newly discovered compound from the soft coral *Gersemia fruticosa*, on the growth and cell cycle progression of various tumour cells in culture. *Steroids*, 1994, **59**, 274–281.
5. Migliuolo, A., Piccialli, V. & Sica, D. Structure elucidation and synthesis of 3 $\beta$ ,6 $\alpha$ -dihydroxy-9-oxo-9,11-seco-5 $\alpha$ -cholest-7-en-11-al, a novel 9,11-secoesterol from the sponge *Spongia officinalis*. *Tetrahedron*, 1991, **47**, 37, 7937–7950.
6. Mori, K., Sakakibara, M., Ichikawa, Y., Ueda, H., Okada, K., Umemura, T., Yabuta, G., Kuwahara, S., Kondo, M., Minobe, M. & Sogabe, A. Synthesis of (22S,23S)-homobrassinolide and brassinolide from stigmaterol. *Tetrahedron*, 1982, **38**, 14, 2099–2109.
7. Barton, D. H. R., Shioiri, T. & Widdowson, D. A. Biosynthesis of terpenes and steroids. Part V. The synthesis of ergosta-5,7,22,24(28)-tetraen-3 $\beta$ -ol, a biosynthetic precursor of ergosterol. *J. Chem. Soc. (C)*, 1971, 1968–1971.
8. Jäälaid, R., Veressinina, J., Järving, I., Pehk, T. & Lille, Ü. Transformation of ergosterol towards 9,11-secoesterols. In *23rd Estonian Chemistry Days: Abstracts of Scientific Conference*. Estonian Chemical Society, Tallinn, 1997, 42.